

Managing 5 part CBC Services

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Automated Cell Counters

3 part vs. 5 part What is the Diff?



3 part vs. 5 part

Cell Counter	3 part	5 part
Differential count	Neutrophils, lymphocytes, mixed	Neutrophils, lymphocytes, monocytes, eosinophils and basophils
Peripheral Smear	Is a must	PS can be made based on validated flag rules
Ease of use and maintenance	Easy with minimum no of reagents and processes	Requires skilled staff adequately trained for operation and maintenance
Cost	Cheap	Relatively Expensive

3 part vs. 5 part



Cell Counter	3 part	5 part
Principle of operation	Impedance based	Impedance based Fluorescence Flowcytometry (Sysmex –XE,XN series, Abott Cell Dyn) Volume Conductivity Scatter (Beckman coulter – LH series) Peroxidase staining (Seimens Advia)
Additional parameters	_	 Reticulocyte Count NRBC Immature platelet fraction Immature granulocytes Additional Scattergrams and flags



Ease of Use

	3 part	5 part
Machine initiation and Software	Easy automated process	Multiple steps. Requires technology savvy personnel.
No of reagents	2 to 3	5-15. Inventory management is critical
End user maintenance	Simple	Complicated and requires training



Cost per test

- 3 part counter
 - Cost of 2-3 reagents per Cycle
 - Sleep mode and startup -shutdown cost
 - Controls
 - Taxes

Approximately Rs. 20-30 per (CBC+Diff) test all inclusive

Details required for Costing



- Average no. of Samples per day for 25 days a week or 300 days a year
- Startup shutdown cycles per year- 300-365

Sr.	Reagent	Pack size	Price / pack	CBC +NRBC	CBC+WDF+NRBC	CBC+WDF+RET+NRBC
No.		(ml) (Rs.) [#]		Cycles / pack	Cycles / pack	Cycles / pack
1	Cell Pack DCL	20000	3500	714	606	488
2	Sulfolyser	3000	20000	6000	6000	6000
3	Lysercell WNR	8000	33000	5333	5333	3200
4	Fluorocell WNR	164	43000	8200	8200	8200
5	Lysercell WDF	8000	32000	0	5333	5333
6	Fluorocell WDF	84	39750	0	4200	4200
7	Cellpack DFL	3000	16000	0	0	2000
8	Fluorocell Ret	24	38000	0	0	1200
9	Fluorocell PLT	24	38000	0	0	0
10	Cellclean	50	7030	0	0	0

- Derive Cost per Cycle
- Derive cost per test
- Add cost of Controls
- Cost for repeats and wastage

Cost per Cycle (CBC +NRBC)	19.66
Cost per Cycle (CBC+ WDF +NRBC)	36.00
Cost per Cycle (CBC+WDF+RET +NRBC)	81.20



Cost per test

5 part counter

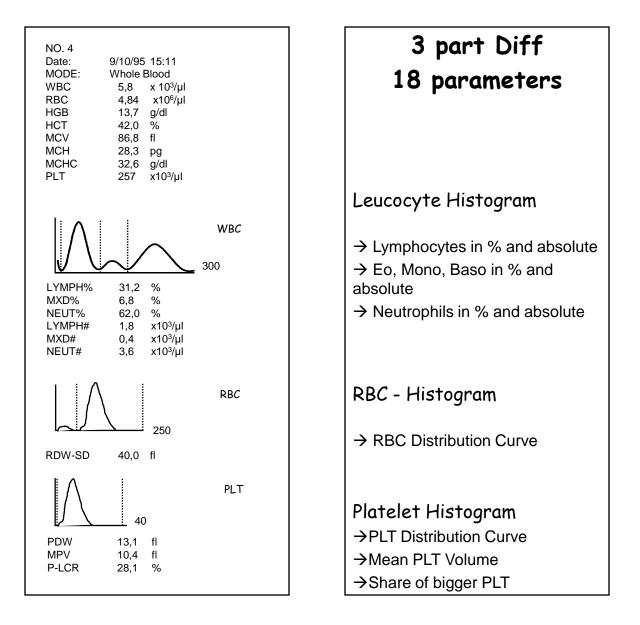
- Cost of reagents per Cycle Complicated process
- Start up and shutdown costs significant addition to costs and is volume dependent
- Controls expensive, short expiry, cost is sample volume dependent
- Taxes
- Hidden costs Always account for Dead volume, background checks, repeats, wastage, EQAS, etc
- Re evaluation of costs after six months 'Consumption based Costing' is a must

Reagent Rental vs. Outright purchase vs. Partial Reagent Rental

Approximately Rs. 50-70 per test (CBC+Diff) all inclusive

3 part Analyser





5 part Analyser

ľ	Main [Graph	WE		BC/PLT	Cumulat	ti۱	/e Q-Flac	s Service HPC		Research(W) Res	earch(R) Rese
Г	Items—			· · ·	WBC Dif			-	Flag(s)	-	DIFF	WBC/BASO
	Item	Data		Unit	Item	Data		Unit	WBC	- 19		MDC/DASC
	WBC	13.34		10^3/uL	NEUT#			10/3/uL	Blasts? Imm Gran?			
	RBC	2.81	_	10^6/uL	LYMPH#			10/3/uL	Left Shift?			
	HGB	7.5	-	g/dL	MONO#			10/3/uL	Atypical Ly?			
	НСТ	22.3	-		EO#			10^3/uL				
	MCV	79.4		fL	BASO#			10/3/uL				
	MCH	26.7		pg	NEUT%	68.6		/0				
	MCHC	33.6		g/dL	LYMPH%	21.2		10			SSC	SSC
	PLT	183		10^3/uL	MONO%	10.0	*	/0			IMI	RET
	RDW-SD			fL	E0%	0.1	*	/0	,	b	2	х Г
	RDW-CV	21.2	+	%	BAS0%	0.1	*	%	RBC/RET	_		-
	PDW			fL	Extende	d Diffe	ere	ential—	Aniso Anemia			
	MPV	10.7		fL	Item	Data		Unit	Aneima			
	P-LCR			% %	IG#	Data		10/3/uL				
	PCT RET%			%	IG%	9.1	*					
	RET#			∽ 10∧6/uL		5.1						
	IRF			%	Item	Data		Unit			DC	SFL
	LFR			%	HPC#			10/3/uL	PLT		RBC	PLT
	MFR			%	Extende	d Paran	net	ers			\land	
	HFR			%	Item	Data		Unit			+1X	X
	NRBC#			10^3/uL	RET-He	Dala						
	NRBC%			/100wBC	IPF		-	pg %			250fL	40fL
								/0				· · · · · · · · · · · · · · · · · · ·
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Principles of 5 part instruments

Impedance and Optical Light Scatter combined with

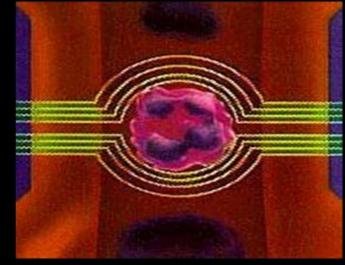
Volume Conductivity Scatter (Beckman coulter – LH series)

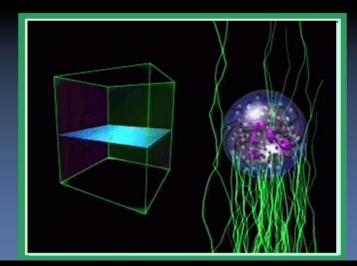
Peroxidase staining (Seimens Advia)

Fluorescence Flowcytometry (Sysmex –XE,XN series)

Beckman Coulter VOLUME MEASUREMENT

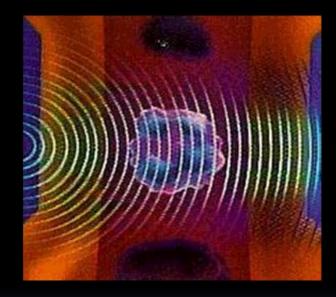
VCS utilises the Coulter Principle of counting and sizing to measure the volume of the cell by using Direct Current (DC) across the two electrode in a flow cell.

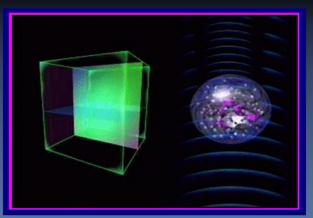




CONDUCTIVITY MEASUREMENT

Cell exposed to RF, the RF energy penetrates into cell and reveal information about its size and internal structure.

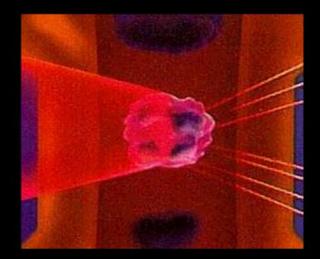


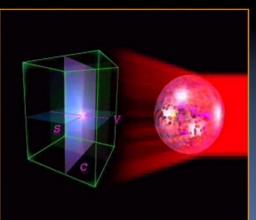


SCATTER MEASUREMENT

As cells are pass in single stream (flow cell) they are struck by laser strike which gets scattered.

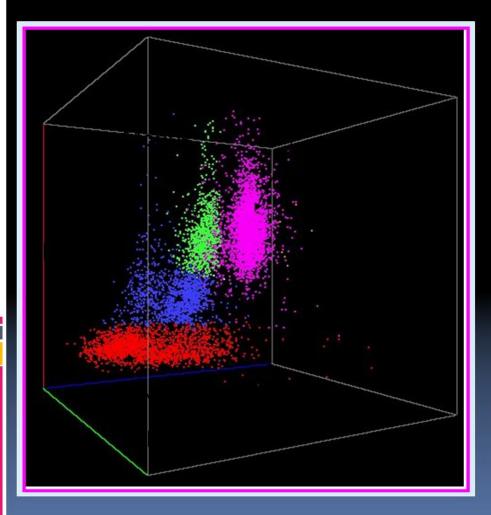
The light scatter at angles between 10 and 70 deg is used by VCS instruments.

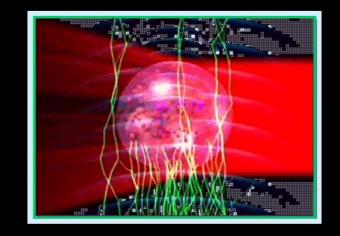


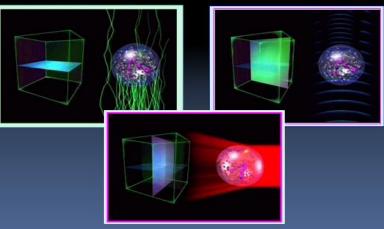


The scattered light gives information about cell surface and granularity

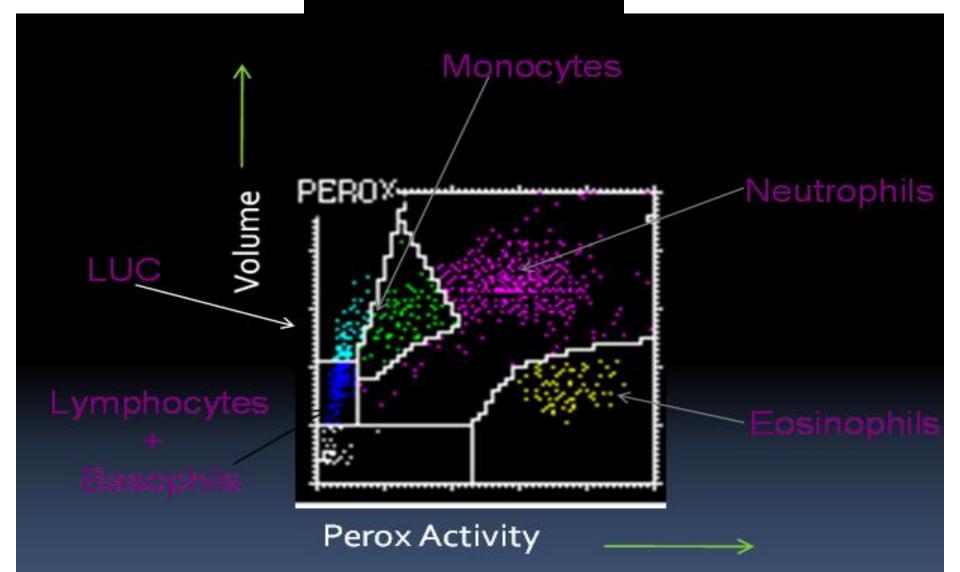
3D Data Analysis







ADVIA TECHNOLOGY

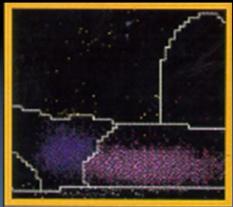


ADVIA TECHNOLOGY

The ADVIA WBC differential is <u>calculated</u> from a 3 step process.

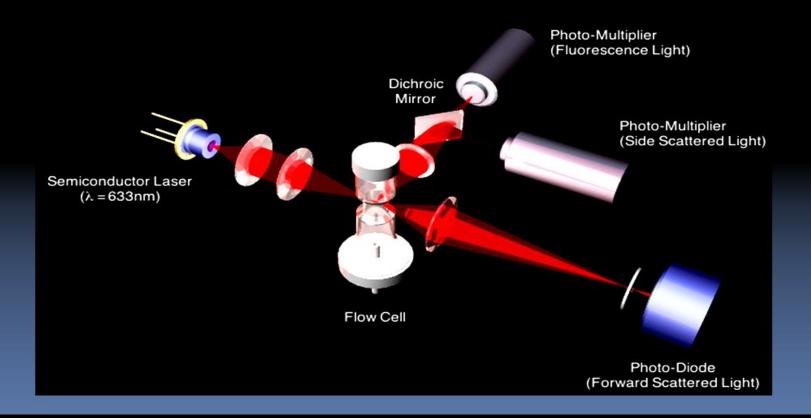
- Cells are stained by peroxidase reagent and analyzed for size and peroxidase stain intensity.
- Cell specific lysis reagents are used to separate basophils from all other white cells.
- Basos are subtracted from the lymph/baso cluster in the perox channel to calculate the lymphs.





Sysmex X-class analyzers-Fluorescence flow cytometry

Optical System



Fluorescence flow cytometry-(light scatter and fluorescent dyes)

Laser Flowcytometry

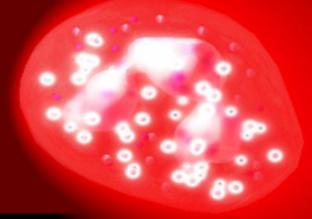
Side Fluorescence Light : RNA/DNA Information



Side Scattered Light : Intenal Cell Structure

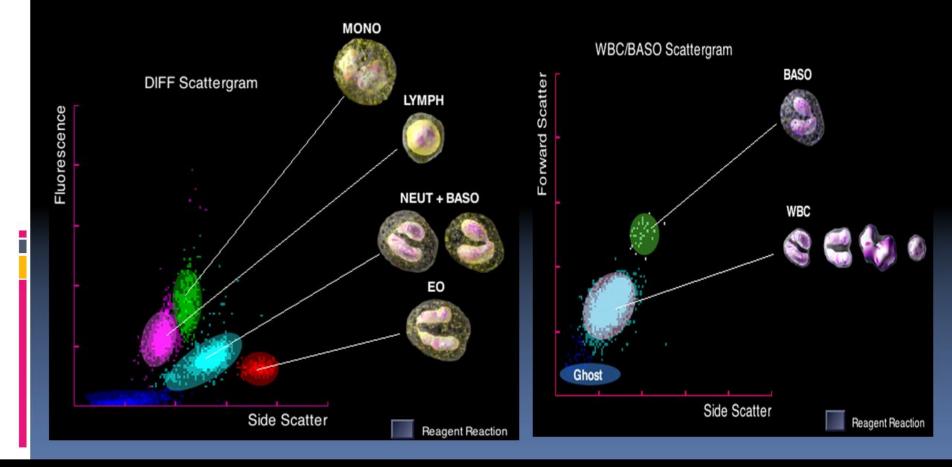
> Forward Scattered Light : Cell Volume Information





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Differential-SFL vs SSc (diff channel) FSc vs SSc (baso channel)





Basic parameters on a CBC Analyzer

- Basic hematological parameters
 - Hb, Hct, RBC Count
 - WBC with Differential (3 part/ 5 part)
 - Platelets
- Derived parameters
 RBC: MCV, MCH, MCHC, RDW
 PLT: MPV, PDW, P-LCR
- Histograms / Scattergrams



Additional parameters

- Principle of measurement
- Clinical relevance and uses
- Limitations

Novel parameter Am J Clin Pathol 2008;130:104-116	Machines	Clinical uses	Limitations
Immature reticulocyte fraction	Sapphire; Pentra 120 DX; LH 750; ADVIA 2120, XE 2100	Classification of anemias; monitoring the efficacy of therapy in nutritional anemia; early identification of marrow regeneration (after bone marrow transplantation or chemotherapy);	Not standardized; reference intervals method-dependent; higher sensitivity in fluorescence-based analyzers
Reticulated platelets	XE 2100	Differential diagnosis for causes of thrombocytopenia	Reduced availability; Lab ranges need to be derived
Immature granulocytes	XE 2100	Diagnosis of bacterial infections	Reduced availability
Nucleated RBCs	Sapphire, Pentra120 DX, LH 750, ADVIA 2120, XE 2100.	Diagnosis of hematologic diseases; prognostic factor in patients from surgery department or undergoing stem cell transplantation; evaluation of the efficacy of transfusion therapy in thalassemic syndromes	Higher performance On fluorescence Based methods
RBC fragments	ADVIA 2120, XE 2100	Diagnosis and monitoring of microangiopathies	Reduced availability; not standardized;
CHr, Ret He	ADVIA 2120, XE 2100	Diagnosis of iron-deficient erythropoiesis	Reduced availability
Hematopoietic Progenitor Cell mode	XE 2100	Surrogate for CD34 stem cell quantitation	Reduced availability, high imprecision



RETICULOCYTE MODE PARAMETERS

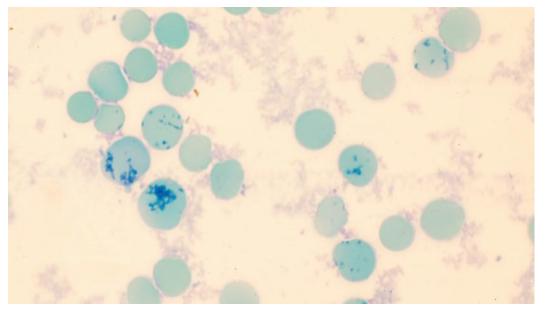
	Test Reticulocyt RPI (Reticu	e Count locyte Production Index)	Result 0.17 0.10	<u>Units</u> %	<u>Status</u> L	Reference Ra 0.20 -	<u>nge</u> 2.50
	Comments :	RPI=Corrected Reticulocyte Col RPI should be used only for adu whereas RPI<2 indicates reduce	It anemic patie	nts. RPI>2 indica	ates significantly	increased hem	atopoiesis
	IRF (Immati	ure Reticulocyte Fraction)	1.20	%	L	2.00 -	16.50
	Comments :	IRF gives an idea about the leas situations the IRF increases bef					
Reticulocyte		iculocyte Hb equivalent) 3C Hb equivalent)	20.80 26.30	pg pg	L	28.70 - 26.00 -	34.10 30.40
Count New Report	Comments :	Ret He provides an idndirect me reduced in patients with Functio normal to high but iron is not del inflammation, cancer patients).	nal Iron Deficie	ncy(FID), FID oc	curswhen réticu	lloendothelial s	
Format	IPF (Immatu	re Platelet Fraction)	6.40	%	Н	0.70 -	4.30
	Comments :	IPF is raised in patients with per low in patients with BM failure. I or stem cell transplant.					

Comments: Ref.-C.Briggs.Int.Jnl>lab.hematol.2009,31,277-97



Manual Reticulocyte Count

- Tedious
- Labour Intensive
- Subjective
- Very High CVs





Automated Reticulocyte Count

PROS

CONS

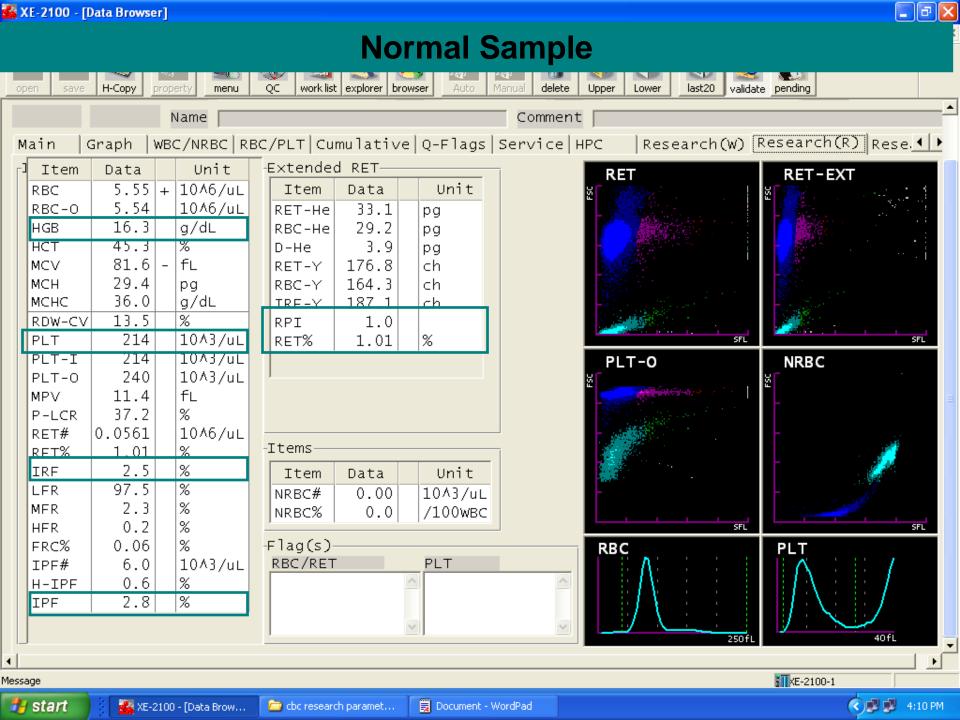
- Rapid
- Reproducible
- Reliable
- Research
 parameters

- Expensive
- Different machines use different dyes and techniques
- Standardisation is difficult
- Reference ranges to be established by every lab

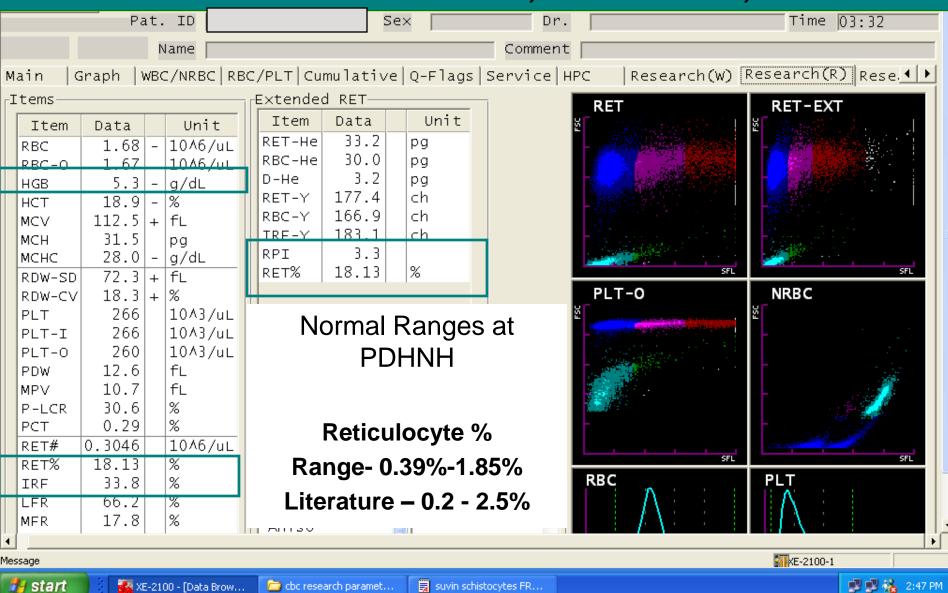


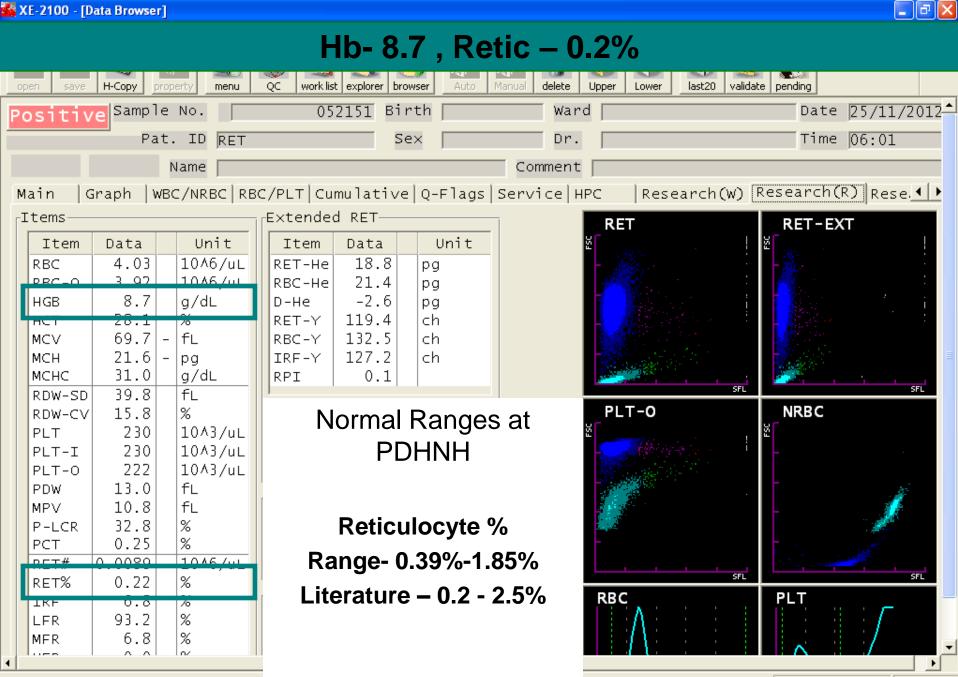
Interpretation of Retic Count

High Retic Count	Low Retic Count
Blood Loss	Nutritional Deficiency- IDA,B12 deficiency
Hemolysis	Aplastic Anemia
Response to therapy	Post Chemo-radiation
Repopulating BM	BM infiltration- benign or malignant diosorders



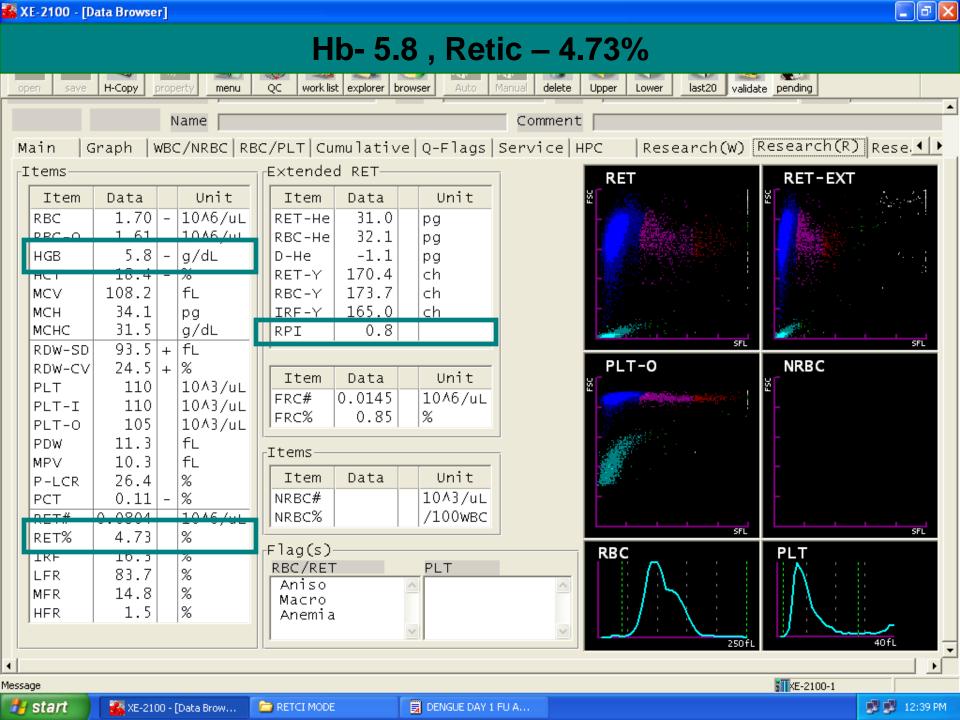
17/M Anemia- Hb-5.3, Retic-18.13%,





Message

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Reticulocyte Production Index-RPI

Reticulocyte Production Index is calculated as follows:

 $\textbf{1}.ReticIndex = ReticCount * \frac{Hematocrit}{NormalHematocrit}$

A value of 45 is usually used as a normal hematocrit.

2. The next step is to correct for the longer life span of prematurely released reticulocytes This relies on a table:

Hematocrit (%)	Retic survival(days) = maturation correction	PB	BM
36-45	1.0	1	3
26-35	1.5	1.5	2.5
16-25	2.0	2	2
15 and below	2.5	2.5	1.5

So, in a person whose reticulocyte count is 5%, hemoglobin 7.5 g/dL, hematocrit 25%,

$$RPI = \frac{ReticIndex}{MaturationCorrection} \rightarrow \mathbf{RPI} = \frac{5 * \frac{25}{45}}{2} = 1.4$$



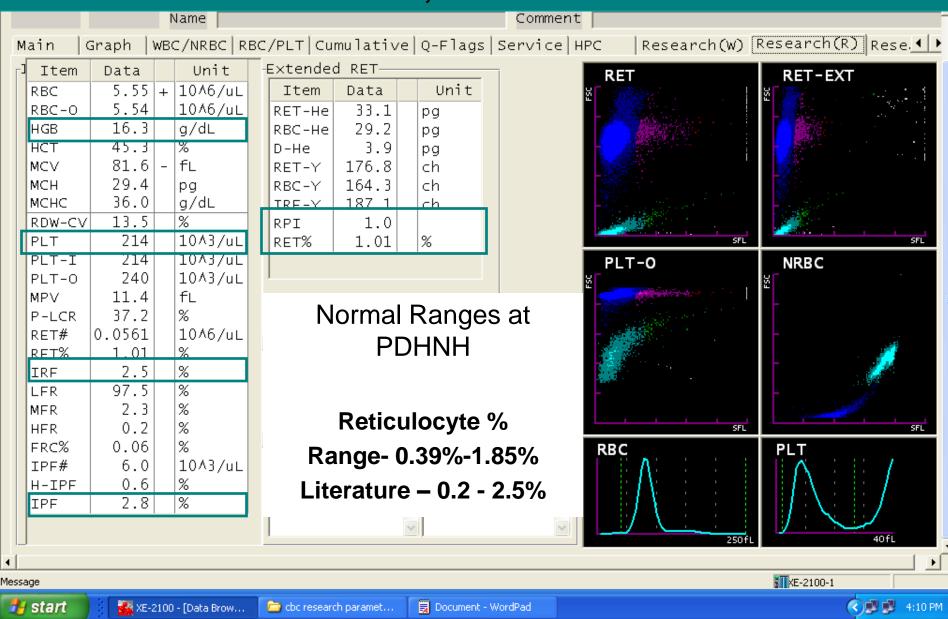
RPI Interpretation

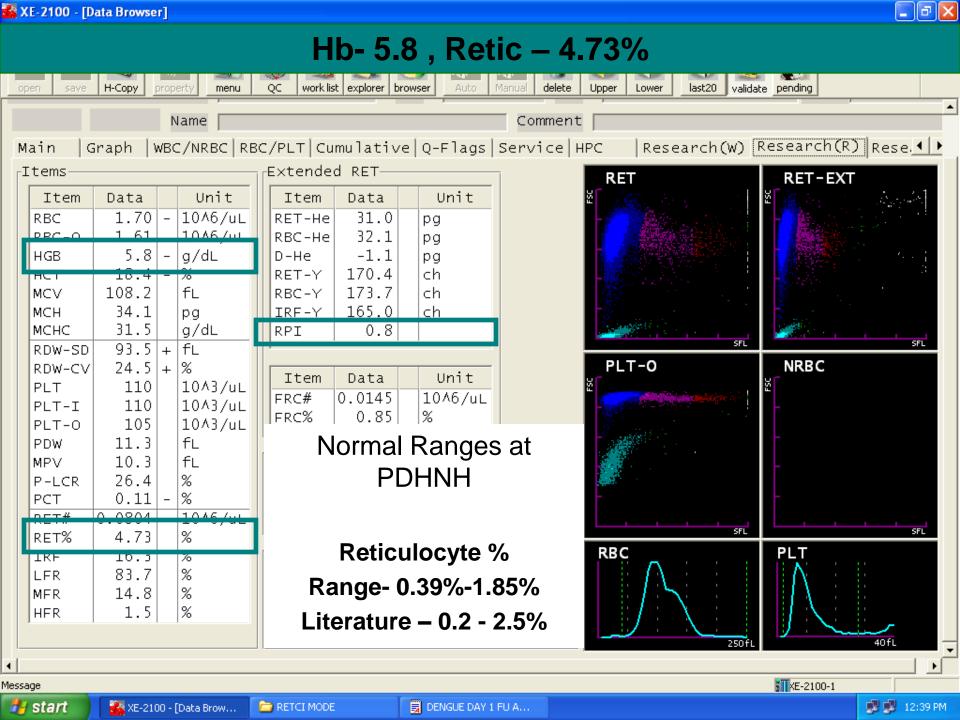
•RPI is used for evaluation only in anemic patients

•RPI < 2 with anemia - Decreased production of reticulocytes and therefore red blood cells.

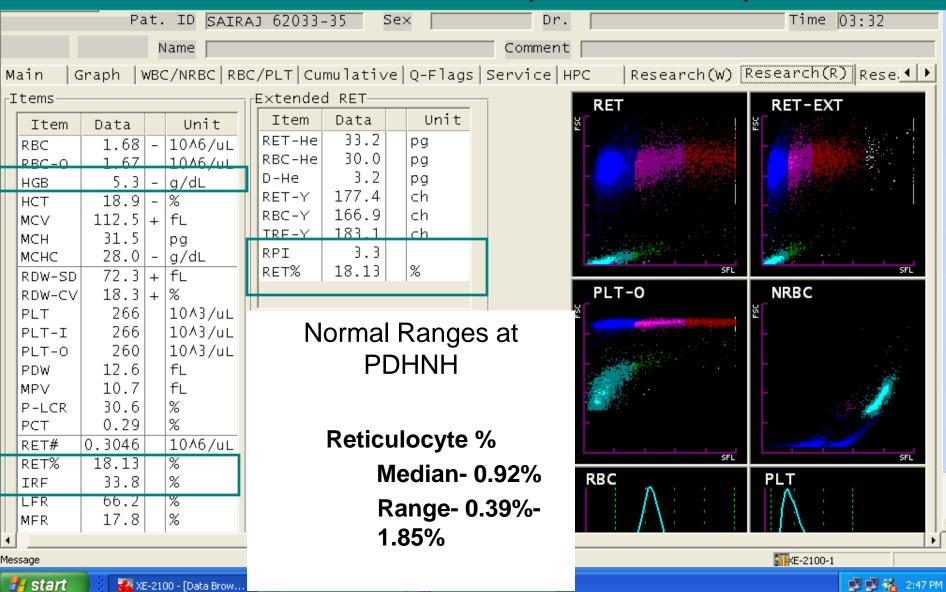
•RPI > 2 with anemia - loss of red blood cells (destruction, bleeding, etc) with increased compensatory production of reticulocytes

Normal Hb-16.3, Normal Platelet Count-214, Normal RPI-1,Normal IRF and IPF





17/M Anemia- Hb-5.3, Retic-18.13%,





Heilmeyer Staging

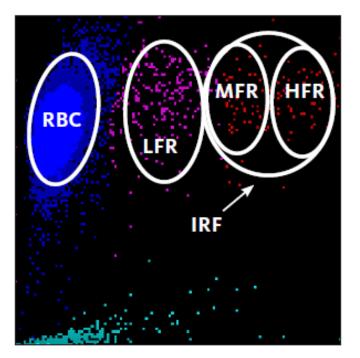
Maturation stages	Morphological	Quantification according		
according to Heilmeyer	description	to Seip (normal %)		
Stage o	Nucleolus			
Stage I	Reticulum consists of dense clots	< 0.1		
Stage II	Loosely arranged reticulum	7.0		
Stage III	Diffusely arranged reticulum	32.0		
Stage IV	Some scattered granulae	61.0		





LFR	MFR	HFR
Low	Medium	High
Fluorescence	Fluorescence	Fluorescence
Reticulocytes	Reticulocytes	Reticulocytes
Little RNA	More RNA	High level of RNA
Mature reticulocytes	Semi-mature reticulocytes	Immature reticulocytes
Reference range:	Reference range:	Reference range:
86.5 - 98.5%	1.5 - 11.3%	0 - 1.4%

Tab. 6 Maturation stages of reticulocytes



IRF is the sum of MFR and HFR, i.e. the immature reticulocytes, and is referred to as the 'reticulocyte maturation index'.

IRF = MFR + HFR

Reference range								
IRF:	f	1.1 – 15.9 %						
	m	1 0 - 12 7 0/						

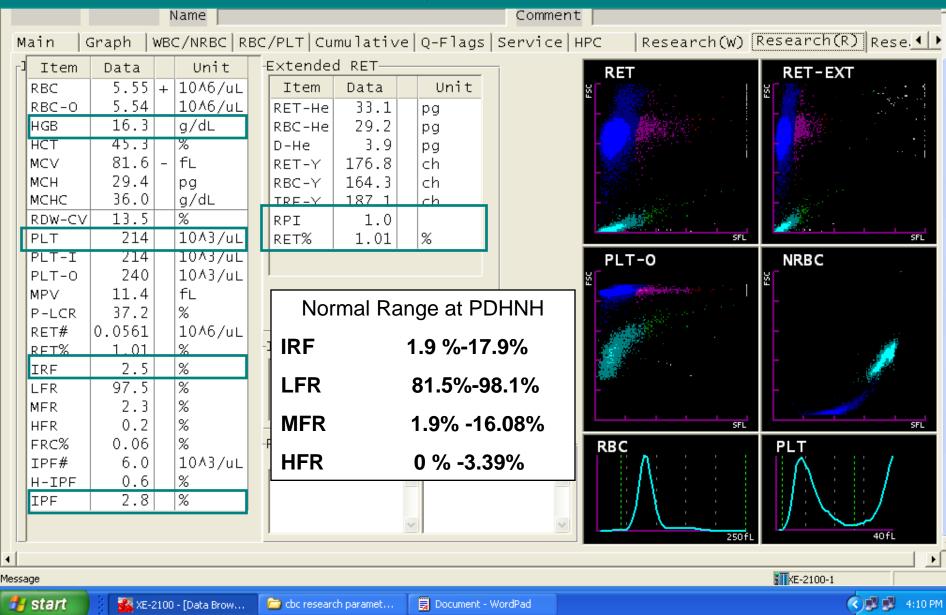
m 1.5 – 13.7 %

In-vitro stability of IRF 6 hours

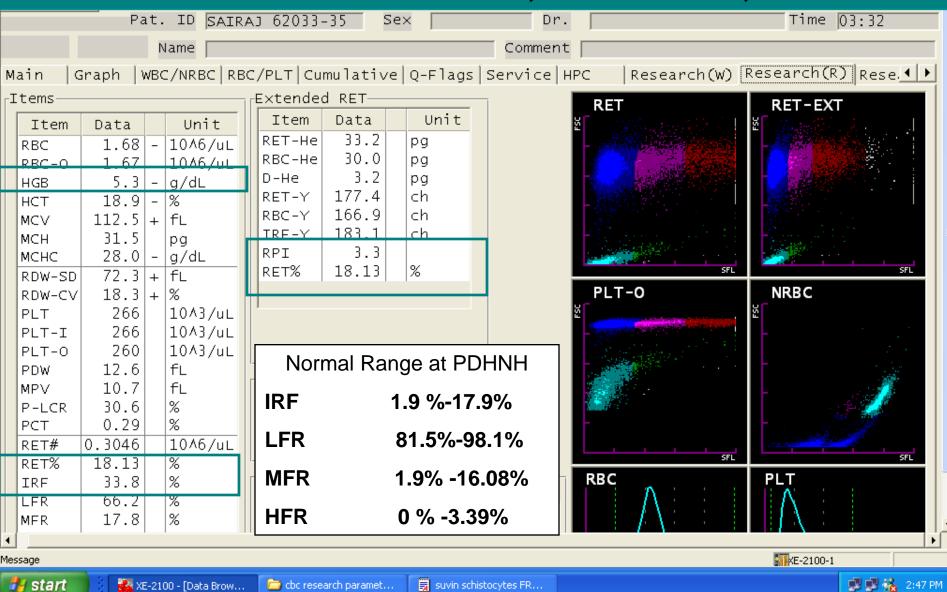
Normal Range at PDHNH					
IRF	1.9 %-17.9%				
LFR	81.5%-98.1%				
MFR	1.9% -16.08%				
HFR	0 % -3.39%				



Normal Hb-16.3, Normal Platelet Count-214, Normal RPI-1,Normal IRF and IPF

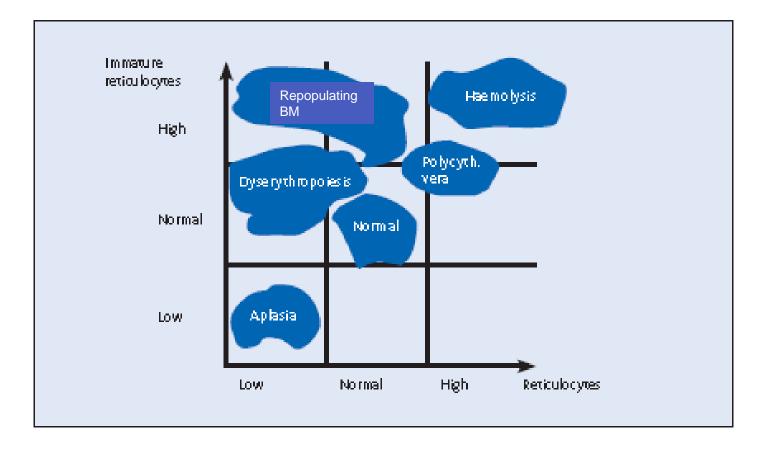


17/M Anemia- Hb-5.3, Retic-18.13%,





Clinical Use of IRF





Indicators of haematopoietic recovery <u>after bone marrow transplantation</u>: the role of reticulocyte measurements.

d'Onofrio G, Tichelli A, Foures C, Theodorsen L.

Universita Cattolica, Roma, Italy.

Abstract

The aim of this project was to study **haematological recovery in patients** following different types of bone marrow transplantation (**BMT**).

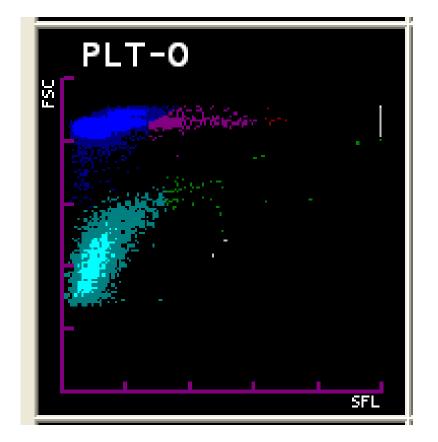
Among 12 different variables, the parameters with the highest specificity or predictive value for monitoring recovery were the **absolute neutrophil count** (ANC) of 0.5 x 10(9)/l, an **absolute reticulocyte count** (RET) above 20 x 10(9)/l **high fluorescent reticulocyte fraction** (HFR) above 5%.

Among these variables, the HFR fraction was the earliest and most sensitive index of engraftment in 79.1% of patients, HFR recovery requiring a median time of 13 days after infusion, in comparison with a median period of 19 and 18 days, respectively, for RET and ANC (P<0.0001).



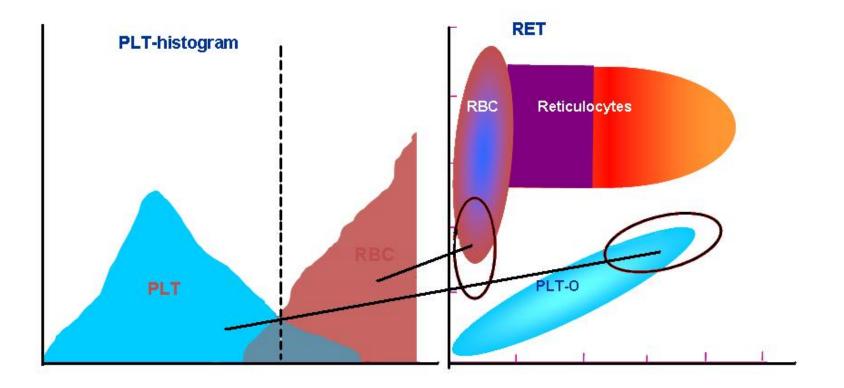
Platelet Parameters

- Optical Platelet Count
- Immature Platelet fraction



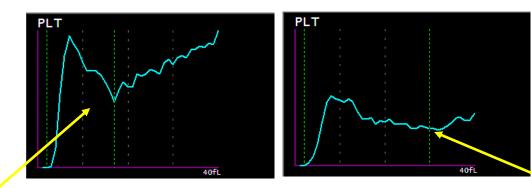


Advantages of Optical Platelet Counting

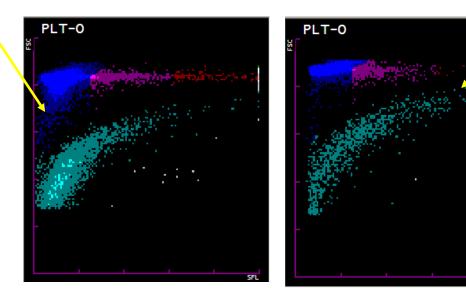


Optical Platelet Enumeration





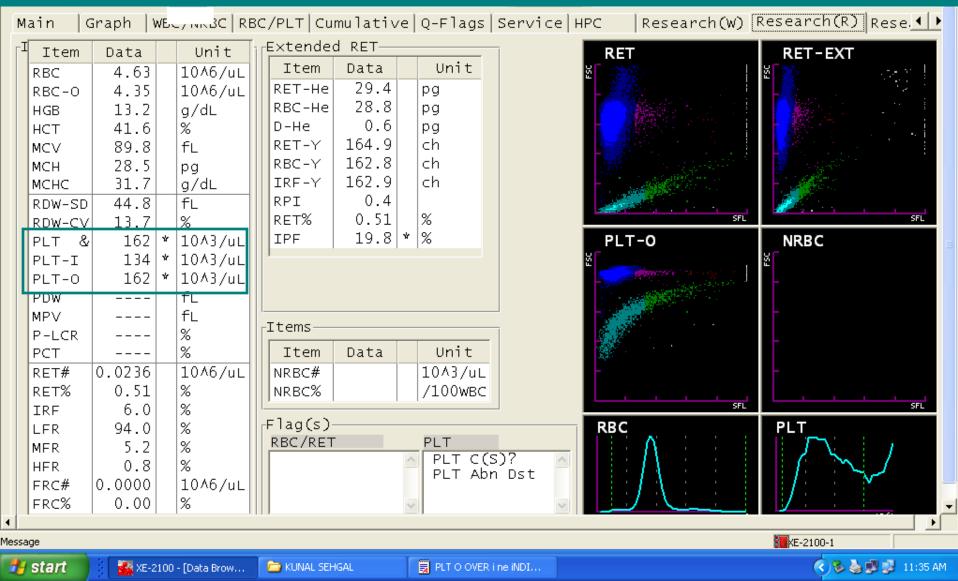
Microcytic RBC



Giant PLT

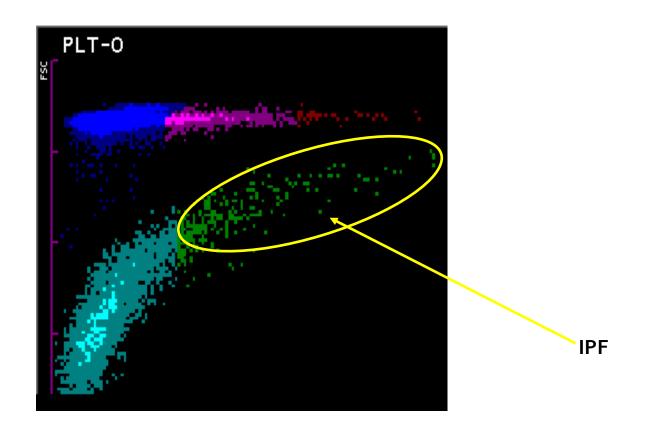
31/F,Blood Donor, East Indian Origin,

Normal Hb and WBC, Impedance Plt- 134, Platelet O –162, Morphologically- Many Giant platelets



Immature Platelet fraction





Immature PLT are identified by its **increase in fluorescence** (more RNA), **FSC is also higher**.

Immature Platelet fraction



1) Pathogenesis of low platelet count:

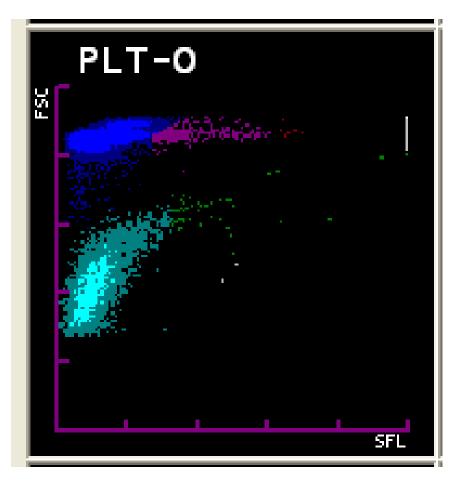
- Increased destruction or usage: Increased IPF
 High IPF suggests an active bone marrow. (e.g., ITP)
- Decreased production: Reduced IPF
 Low IPF suggests depressed bone marrow function

2) Bone Marrow regeneration :

IPF - First indicator of bone marrow regeneration

FDA approved

IPF - Normal Range



IPF – Various studies have shown Normal range as 0.5 to 5.2%, 1.1 to 6.1%, 0.5 to 3.2 %

Normal Ranges Derived at Hinduja Hospital

• IPF - 0.7-4.3%



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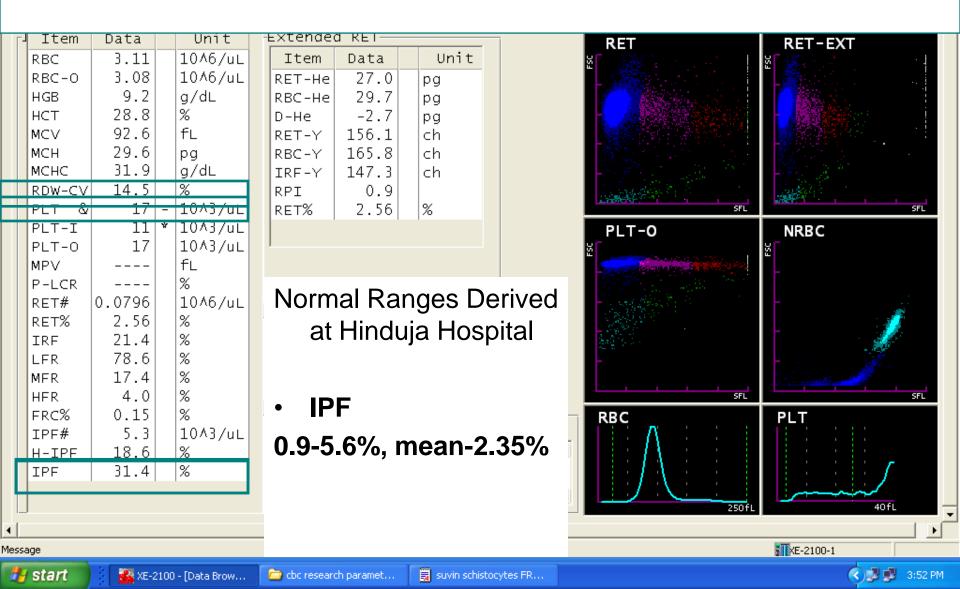


Normal Hb-16.3, Normal Platelet Count-214, Normal RPI-1,Normal IRF and IPF

	Name Comment									
N	1ain	Graph	WBC/NRBC	RBC/PLT Cu		e Q-Flags	Service	HPC	Research(W)	Research(R) Rese. • •
Ē] Item	Data	Unit	-Extende	d RET			RET		RET-EXT
	RBC	5.55	+ 10^6/uL	Item	Data	Unit		2 🔐	\$	× 1
	RBC-0	5.54	10^6/uL	_ RET-He	33.1	pg				
	HGB	16.3	g/dL	RBC-He	29.2	pg				
	НСТ	45.3		D-He	3.9	pg				
	MCV	81.6	- fL	RET-Y	176.8	ch				
	MCH	29.4	pg	RBC-Y	164.3	ch				
	MCHC	36.0	g/dL	TRF-Y	187 1	ch				
	RDW-C			RPI	1.0					
	PLT	214	· · ·	- RET%	1.01	%			SFL	SFL
	PLT-I	214	'					PLT	-0	NRBC
	PLT-0	240	10^3/uL fL	-				FSC	Contraction of the local sectors of the local secto	ы Парала Пара Пар
	MPV P-LCR	37.2				_		10.5		
	RET#	0.0561	/~ 10^6/uL	Norm	nal Rai	nges De	erived			
	RET%	1.01	10,007ut	-		•			53 C	
	IRF	2.5	%		ΠΙΠάι	ija Hosp	Jilai		11 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1	
	LFR	97.5		-				2000 C	-	
	MFR	2.3	%							
	HFR	0.2	%			7- 4.3%			SFL	SFL
	FRC%	0.06	%		F • V.	/- 4.3 /0)	RBC		PLT
	IPF#	6.0	10^3/uL	-					Λ	$\square \square \land \square \square \square \square$
	H-IPF	0.6								[[]] X := []
	IPF	2.8	%							
	J								250fL	40fL
•										_
Mess	age									₩E-2100-1
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15/f with petechial rash and thrombocytopenia -17x10³,

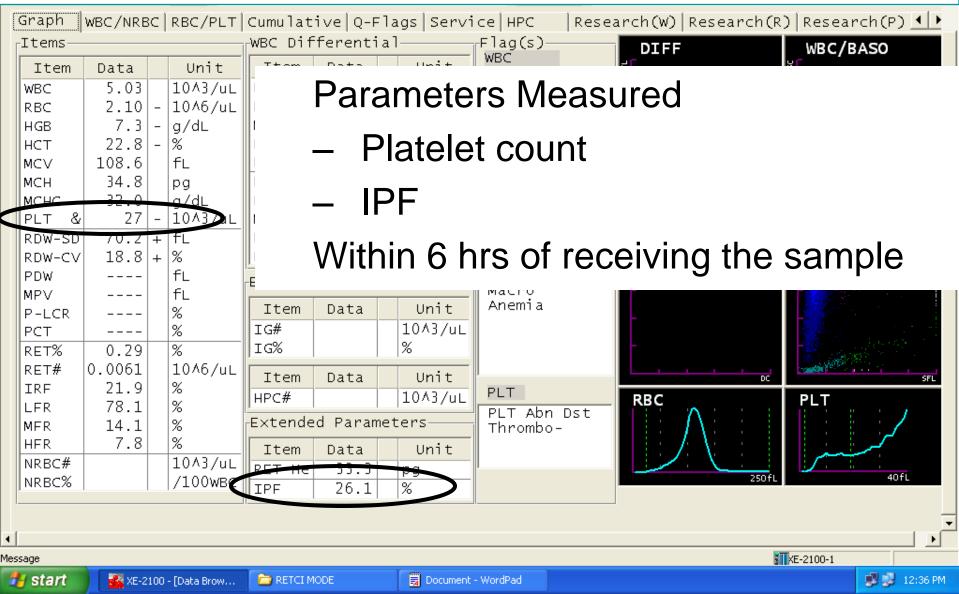
IPF-31.4%





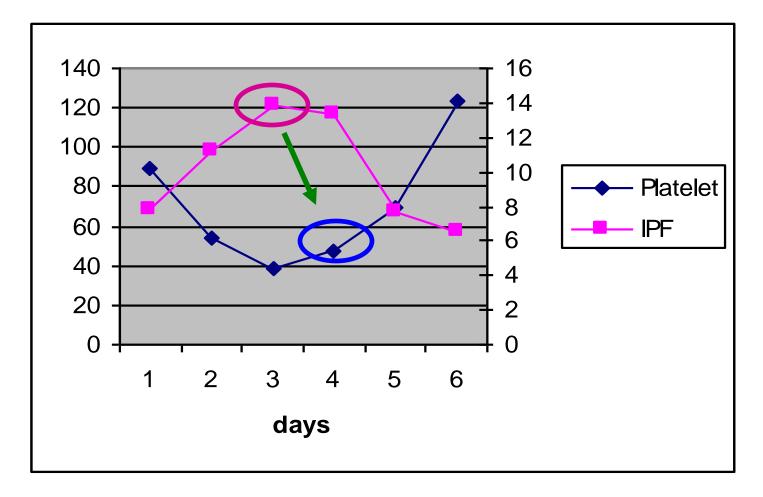
Dengue NS1 Ag positive – KHAR HNH

Platelet - 27000, IPF - 26.1%





DAYS	1	2	3	4	5	6
PLATELETS	89	54	30	47	70	123
IPF	7.8	11.2	13.9	13.3	7.7	6.5





IPF in Dengue

- IPF IS A POTENTIAL TOOL FOR PREDICTING PLATELET RECOVERY IN DENGUE PATIENTS HAVING THROMBOCYTOPENIA.
- A SINGLE VALUE OF >10% IS INDICATIVE OF PLATELET RECOVERY WITHIN 24-48 HRS

Summary



- Many new parameters have been added to the conventional CBC.
- Increased sensitivity and precision (e.g., IGs, nRBCs).
- Parameters like Ret-He do not have another comparable/ manual method, making such a parameter invaluable
- Improved Turn around Time

Summary



 IPF gives insight into the pathogenesis of thrombocytopenia. Can help avoid unnecessary BM examination and platelet transfusions.

 Newer parameters add a lot of extra information to the standard CBC, which may translate into better patient care.

 Novel Parameters still need to be standardised across instruments and labs should make their own normal ranges before using them

